

COLLECTION INSTRUCTION FOR CLIENTS

All specimens:

All specimens **must** be accompanied by a requisition slip with the following patient information:

- Patient's name
- Patient's date of birth and sex
- Physician's name
- Insurance information
- Body site of specimen
- Any pertinent clinical history
- Pre-op diagnosis
- Collection date and time

All specimens should be submitted in containers with two unique patient identifiers and proper fixative hazard label.

Routine specimens:

As soon as possible after removal, place in 10% neutral buffered formalin container, labeled with at least 2 patient identifiers. To ensure best fixation results, the tissue should be in an adequate amount of fixative (10-20 times the tissue volume).

Place in a properly labeled container, containing formalin. Place container in an individual sealable biohazard bag, if possible, with the accompanying requisition slip in the outside pocket. If specimen is too large to fit in bag, attach requisition to the container with tape. One requisition can accompany multiple specimens, from the same patient, but each specimen should be placed in its own container and listed separately on the requisition.

Contact Lab to arrange pick up of the specimen(s).

Direct immunofluorescent studies (DIF):

Place specimen in a properly labeled container containing Michel's Fixative. Keep stored in refrigerator until picked up.

Breast specimens for Estrogen and Progesterone receptors and Her2/Neu (c-erB-2) studies:

Breast tissue (biopsy, mastectomy, mammoplasty) should be placed in 10% neutral buffered formalin within one hour of removal. Both time of removal of the tissue and the time of immersion in fixative must be recorded on the requisition for valid results. The

tissue should not have been in formalin for more than 72 hours. Call lab if coordination is needed to accommodate holidays and/or weekends.

Uric acid crystals:

Specimen should be submitted in 100% ethyl alcohol, not formalin.

Non-gyn slides:

Slides submitted must have the patient's name and body site written on the slides and can be fixed or air-dried, but must specify which. Slides can be "wet fixed" by submerging freshly prepared smears in an alcohol fixative such as, 95% Ethanol, an Ether alcohol mixture, 100% Methanol, 80% Propanol and Isopropanol, or Denatured alcohol, for a minimum of 15 minutes. Alternatively, slides can be fixed using a coating fixative such as Carbowax or Diaphine fixative spray.

Gross only examinations:

Can be submitted with or without fixative, but if submitted with tissue, it must contain formalin.

Other:

Tissue that cannot be processed immediately or requires special procedures to be performed on tissue prior to fixation, should be refrigerated if the laboratory is closed for the day. The physician office should notify Dominion Pathology Laboratories what studies they require and the technician or pathologist will give direction for the handling of the specimen.

If you have any questions regarding specimen collection, please contact the laboratory at 757-664-7901 for further instructions.